

This is a self-archived – parallel published version of an original article. This version may differ from the original in pagination and typographic details. When using please cite the original.

AUTHOR Anna L. Ruotsalainen, Miia Kauppinen, Piippa R. Wäli, Kari Saikkonen, Marjo Helander, Juha Tuomi,

TITLE Dark septate endophytes: mutualism from by-products?

YEAR 2021, 27th October.

DOI <https://doi.org/10.1016/j.tplants.2021.10.001>

VERSION Author's accepted manuscript

COPYRIGHT License: [CC BY NC ND](#)

T

CITATION Anna L. Ruotsalainen, Miia Kauppinen, Piippa R. Wäli, Kari Saikkonen, Marjo Helander, Juha Tuomi.
Dark septate endophytes: mutualism from by-products?,
Trends in Plant Science. 2021,
ISSN 1360-1385,
<https://doi.org/10.1016/j.tplants.2021.10.001>
(<https://www.sciencedirect.com/science/article/pii/S1360138521002764>)

1 **Dark septate endophytes: mutualism from by-products?**

2 **Ruotsalainen, A.L.^{1*}, Kauppinen, M.², Wäli, P.R.^{1,3}, Saikkonen, K.², Helander, M.⁴, Tuomi, J.⁵**

3 ¹Department of Ecology and Genetics, POB 3000, FI-90014 University of Oulu, Finland

4 ²Biodiversity Unit, FI-20014 University of Turku, Finland

5 ³Natural Resources Institute Finland (Luke), Ounasjoentie 6, FI-96200 Rovaniemi, Finland

6 ⁴Department of Biology, University of Turku, FI-20014 Turku, Finland

7 ⁵Meritie 43, FI-29900 Merikarvia, Finland

8

9 Helander, M. ORCID 0000-0002-9759-4321

10 Saikkonen, K. ORCID 0000-0001-5203-9984

11

12 Wäli, P.R. ORCID 0000-0002-2484-7455

13 *Correspondence: annu.ruotsalainen@oulu.fi (A.L. Ruotsalainen) ORCID 0000-0001-6621-0375

14 **Keywords**

15 cost-benefit, evolution, nutrient uptake, plant-fungal interactions, root-associated fungi, symbiosis

16

17 **Glossary**

18 **Biotrophy:** organism feeding on other organism.

19 **By-product mutualism*:** mutually beneficial interaction between individual organisms equipped
20 with traits that primarily benefit the bearer and benefit the other individual only as a side effect.

21 **Mutualistic association:** mutually beneficial interactions between two individual organisms.

22 **Mycorrhizal symbiosis:** symbiosis between plant roots and fungi, in which the fungus facilitates
23 nutrient uptake from soil into the plant and gets carbon in return.

24 **Pseudoreciprocity*:** organisms's investment on itself benefits another organism as a by-product.
25 Insect mating gifts, where the male investment on their own reproduction benefits female mating
26 behaviour is an example of pseudoreciprocity.

27 **Reciprocal investments*:** continuous reciprocal investments between individual organisms.
28 Investment can come in many forms, for example (1) improving physical access to partners and
29 their resources/services, (2) improving/manipulation of the partners ability to provide beneficial
30 services/resources, and (3) improving ability to effectively utilize the received services/resources
31 for own survival and reproduction.

32 **Saprotrophic capacity:** ability to feed dead organic material.

33 **Symbiosis:** living together. A close interaction between two organisms. The outcome of the
34 interaction can be positive (mutualism), neutral (commensalism) or negative (parasitism).

35 *Modified after [35,41]

36

1 **Abstract**

2 Plant roots are abundantly colonized by dark septate endophytic (DSE) fungi in virtually all
3 ecosystems. DSE fungi are functionally heterogeneous and their relationships with plants range from
4 antagonistic to mutualistic. Here, we consider the role of by-product benefits in DSE and other root-
5 fungal symbioses. We compared host investments against symbiont-derived benefits for the host plant
6 and categorised these benefits as by-products or benefits requiring reciprocal investment from the
7 host. By-product benefits may provide the variability required for the evolution of invested
8 mutualisms between the host and symbiont. We suggest that DSE could be considered as “by-product
9 mutualist transitional phase” in the evolution of cooperative mycorrhizal symbionts from
10 saprotrophic fungi.

11

12 **DSE - root fungal symbionts between mycorrhizal and saprotrophic habit**

13 Dark septate endophytic fungi (DSE, [1] (Box 1) colonize plant roots in most taxonomic groups in
14 all major biomes of the world [2]. Although DSE colonization has been shown to be able to improve
15 growth and nutrition of the host plant [3,4] and thus resemble **mycorrhizal symbiosis** (Glossary), the
16 nature of DSE symbiosis – whether beneficial or harmful for the host plant- has remained largely
17 unknown. Here, we define DSE as fungi that colonize living plant roots by melanized septate
18 (ascomycetous) hyphae and sometimes microsclerotia (Box 1, Figures 1 and 2). DSE is an
19 unambiguous form group and may represent several orders within ascomycetous fungi [5]. Recent
20 studies have also found DSE to be characterized with a marked proportion of saprotrophic genes in
21 their genomes [6-8]. High-throughput sequencing of soil has revealed an abundance of DSE fungi in
22 plant rhizospheres (e.g., [9-12]). Interest towards applications in plant production [13-15],
23 phytoremediation [16-23] and in carbon sequestration into soil [24]
24 (<https://www.theland.com.au/story/5344438/soil-survival-benefits-from-a-fungi/>) would benefit
25 from improved understanding of the biology of DSE symbiosis.

26 DSE fungi thus colonize roots of healthy plants by forming both superficial and intraradical fungal
27 hyphae (Figures 1 and 2) and by forming intraradical microsclerotia (Figure 2). Bidirectional
28 translocation of carbon and nutrients between host plants and root-associated fungi is the core
29 definition of mycorrhizal symbiosis [25] and therefore, is also of major interest when studying
30 nutritional benefits of DSE for plants. The transfer of carbon from the host plant to DSE fungi has

1 been detected to take place [10,26] although it is not clear whether all the transfer is due to intraradical
2 fungal colonization [27]. Improved nitrogen acquisition of the host plant, which is often reported
3 resulting from DSE colonization, is neither necessarily directly associated with DSE colonization in
4 plant roots [27]. In addition, the ecological significance of carbon and nutrient translocation between
5 the host plant and DSE fungi - if it takes place - is not well understood because plant responses to
6 DSE fungal colonization are context dependent and vary from negative to positive (e. g, [3,4,28,29]).
7 Lack of knowledge of taxonomy and function in relation to morphological definitions is typical in
8 the research field of mycorrhizal and other root-associated fungi.

9 Here, we focus on eco-evolutionary evidence indicating that DSE fungi have properties from both
10 mycorrhizal and saprotrophic fungi [7,8,10,30,31]. Similar to mycorrhizae, DSE fungi form close
11 associations with plants but, similar to soil saprobes, they may also be independent of their host plant
12 because of **saprotrophic capacity**. Furthermore, in DSE symbioses, benefits and costs may not be
13 limited to nutrient and carbon trade to the same extent as in mycorrhizal symbioses (see e.g., [29]).
14 Therefore, theories of mycorrhizal symbioses based on reciprocal investments [32] and on the theory
15 of biological markets [33,34] may not be directly applicable to DSE. Instead, soil properties and
16 resource pools related to the plant-soil interface may play a more significant role. In this opinion, we
17 propose that DSE symbiosis could be better understood by considering **by-product benefits** [35,36]
18 and briefly discuss DSE symbiosis in relation to the evolution of mycorrhizal symbioses.

19

20 **Benefits of mutualism**

21 *Benefits of mutualism*

22 Connor [35] classified benefits of **mutualistic associations** into three categories: (i) by-product
23 benefits, (ii) invested benefits and (iii) purloined (i.e., stolen) benefits. By-product benefits are traits
24 or other attributes of an organism that incidentally benefit the other organism. In by-product
25 mutualism one organism receives benefits that another organism produces as a by-product of its self-
26 serving traits [35,37], such as in the case of certain micro-organisms that utilize the metabolic waste
27 products of their hosts [37,38]. By-product mutualism has been considered an important step and one
28 explanation for the evolution of cooperation [35,36,39-41] (see also Harcombe et al. [38] for
29 empirical results in an experimental bacterial system). In invested benefits one partner actively invests
30 in the other. The benefit of the interaction is then considered to exceed the cost of the investment
31 [35]. In reciprocal interactions both partners invest and benefit from the **symbiosis**. Mycorrhizal

1 symbiosis is often considered a classic example of a mutualistic interaction based on **reciprocal**
2 **investments** and benefits i.e., interspecific transfer of nutrients and carbon between the host and fungi
3 [25,32]. Benefits and reciprocal investments are, however, only rarely symmetrical in nature [42].
4 For example, similarly to all other biological interactions, mycorrhizae-host interactions are based on
5 reciprocal exploitation [43] and despite generalizable expressions (such as “exchange” and “trade”)
6 mycorrhizal symbioses include diverse dynamics [44]. In addition, mycorrhizae have also been
7 regarded as an example of **pseudoreciprocity** where the investment from a host plant into root growth
8 increases the availability of new root tips for mycorrhizal colonization, which can be seen as a by-
9 product benefit for the plant [41]. The extreme case are purloined benefits, where symbionts also
10 exploit (i.e, steal) resources that were intended to increase the fitness of the plant partner in addition
11 to by-products. Despite stealing of resources by the symbiont (parasite), the symbiosis may be
12 mutualistic if the parasite produces other benefits - either by-products or invested - for the host plant
13 that exceed the costs of purloined benefits [35].

14
15

16 *Window for by-product benefits*

17 As above stated, in plant-root fungal symbiosis by-products could include resources that the host
18 plant has an excess of or are waste products of the plant. In particular, plants may have excess carbon
19 products and these may enter the soil via many routes (respiration, plant litter, secretion of organic
20 acids and dead root biomass) [45-48] (Figure 3).

21

22 Although the evolution of plant-microbial symbioses can be assumed to be driven by competition for
23 resources between organisms, selection may favor properties that also benefit the other organism. If
24 by-product benefits are present in plant-fungal symbiosis, the optimal investment of the plant to the
25 symbiont may decrease even to the theoretical level where the optimal (best) investment is zero and
26 the plant exploits by-product benefits without investing in the symbiont (Figure 4, highest curve).
27 Contrary to that: when by-product benefits do not exist at all, it may be optimal for plants to either
28 have no investment at all or, alternatively, a relatively high investment into the symbiont is required
29 for symbiosis to be profitable for the host (Figure 4, lowest curve). In cases where the by-product
30 benefits of symbionts is not in either of the previously mentioned extremes, we suggest that it is best
31 for the plant to either not invest at all (i.e. to obtain lesser, but still positive, by-benefits from the

1 symbiont) or to invest in the symbiont, but a lower level of investment is optimal than in the situation
2 where no by-product benefits exist at all and all benefits are only gained via investment (Figure 4).

3
4 Based on this schematic presentation, the presence of by-product benefits may decrease the threshold
5 of selection for costly traits in plants and increase their dependency on the microbial partner, such as
6 DSE. It may be especially important if, in the absence of by-product benefits, investment costs are
7 higher than the corresponding benefits for low investment levels (Figure 4, lowest curve). By-product
8 benefits can thus help plants to overcome this initial bottleneck. Because our presentation only
9 includes treatment from “plant’s view”, it is important to note that as far as the obtained benefits for
10 the host plant require costly investments from the symbionts (for example, altered physiological
11 functions or structural investments) the joint evolution of both of them will determine the outcome
12 (i.e, whether the local plant optima in the can be reached or not).

13 14 15 *DSE fungi and by-product benefits*

16 DSE fungi have a wide variety of enzymes for organic matter degradation [30,49,50] and therefore
17 they resemble free-living soil saprotrophs. DSE fungi have also been found to have a positive impact
18 on plant growth in the presence of organic nutrients [27,51,52] and the colonization of plant roots by
19 DSE fungi in the field correlates positively with the amount of organic matter in soil [53-57].
20 Intraradical colonization of DSE however suggests a special, differentiated relationship with host
21 plants because root colonizing fungi have to cross physical and chemical barriers during entering the
22 root and be able to tolerate conditions inside the roots. Root colonizing fungi thus have to have the
23 ability for a complex cross-talk with the host [58,59]. Intraradical colonization also increases
24 opportunities for close interactions between hosts and fungal symbionts, for example in carbon and
25 nutrient translocation or other potentially beneficial impacts, such as hormonal signalling [60]. By
26 having both the saprotrophic capacity for organic matter degradation in the soil and the capability of
27 colonizing roots, DSE fungi could therefore fit well into the category of by-product mutualists which
28 enhance the performance and fitness of their host plants by providing benefits, but not requiring major
29 investments from the host. In addition, among Pleosporales, which is an order including many DSE
30 fungi, a transfer from saprotrophic to hemibiotrophic and **biotrophic** states during evolution has been
31 suggested [61].

32 33 **Dark septate endophytes – mutualism from by-products?**

1 Evolution in the fungal tree of life comprises a spectrum of symbiotic (mycorrhizal) and saprotrophic
2 lifestyles largely arising from ancestral features of fungi, such as hyphal cell structure, hyphal growth
3 embedded into substrate, extrahyphal enzymes and symbiosis with photosynthesizing organisms.
4 Although evolution of lifestyles rather consists of continuums than “man-made” categories, we
5 compared key characteristics of root-associated and rhizosphere fungal groups to contrast the general
6 differences between mycorrhizal, DSE and saprotrophic fungi (Table 1). This simple comparison
7 shows that DSE symbiosis resembles free-living saprotrophic fungi more than mycorrhizal fungi.
8 When paying specific attention to symbiont-derived benefits for the host and specificity of the
9 association between the host and the symbiont (Table 1), DSE symbiosis can be merely considered
10 as an intermediate, transitional form. Thus, DSE fungi are more beneficial for their hosts and have
11 higher host-specificity than free saprotrophs, but they are less beneficial and have lower host-
12 specificity than mycorrhiza-forming fungi (especially Glomeromycota, the ancestral form of plant-
13 fungal symbiosis, see Jumpponen et al. [2] for DSE and Smith and Read [25] for Glomeromycota).
14 In line with this, different fungal phyla are involved in divergent functional roles (Table 1) [62].

15

16 By-product benefits may also play a role in evolution of mycorrhizal symbioses in general [41,63-
17 66]. For example, Martino et al. [59], based on a genomic analysis, showed that development from
18 an endophytic state has taken place during the evolution of ericoid mycorrhizal symbiosis. Ericoid
19 mycorrhizal fungi resemble DSE fungi: they both have developed enzymatic capacity for organic
20 matter degradation and they both occur as non-mycorrhizal endophytes in the roots of a wide variety
21 of plant groups. However, in contrast to DSE, ericoid mycorrhizal fungi also form a highly specialized
22 and functionally well-characterized mycorrhizal symbiosis with ericaceous plants [24]. Similarities
23 in the life strategies between DSE fungi and ericoid mycorrhizal fungi give support for hypotheses
24 about a relatively recent transition between symbiotic and saprotrophic growth habits among the
25 fungal lineages. There are also other mycorrhizal groups which may be less known but have well-
26 developed saprotrophic capacity, such as orchid [25] and sebacinoid mycorrhiza [65,66]. It is possible
27 that our treatment/discussion is applicable to this kind of mycorrhizal fungi as well.

28

29 Similar to other biotic interactions, fungal symbioses are dynamic and context dependent continuums
30 of interactions with host plants [43]. Consequently, mutual dependency between the fungus and the
31 host plant may be less likely to evolve in heterogeneous environments where the benefits of the
32 cooperation vary [64,67]. Chamberlain et al. compared interaction types (competition,
33 commensalism, mutualism) and showed that mutualism was most likely to change to neutral or
34 antagonistic according to the context [68]. More recent analyses based on phylogenetic data indicated

1 that evolutionary history would better explain large-scale mutualism breakdown/speciation events
2 than context, in particular when nutritional-type symbioses are considered [67, see also 69]. It could
3 be that if the amount of by-products varies in space and time it could favor organisms that are flexible
4 in investment strategies and are able to change the amount of investments according to the availability
5 of by-products and the range of symbiotic partners available. This could well be the case in most
6 terrestrial ecosystems. By-product benefits should therefore be seen as a potential step towards
7 mutualistic relationships in the evolution of plant-fungal interactions.

8

9 **Concluding remarks**

10 We conclude that the presence of by-product benefits may increase options for mutualism to
11 evolve. In ecological contexts, by-product benefits may lead positive association of species purely
12 due to improved local population growth rates without particular adaptations to fortificate the positive
13 reciprocal effects. The first evolutionary steps towards the increased dependence might involve
14 costly adaptations (investments) to (i) improve physical access to partners and their
15 resources/services or (ii) improve the ability to effectively utilize the received services/resources for
16 own survival and reproduction. These adaptations may not require reciprocal investments but may
17 eventually lead to the dependency on the presence of the partner and eventually to the coevolution of
18 the interacting species. Secondly, the costly investments/adaptations may specifically involve
19 improvement/manipulation of the partners ability to provide beneficial services/resources in quantity
20 and/or quality above the level of by-product benefits. Our schematic model outlines some
21 hypothetical possibilities for the shape of benefit curves for such costly investments in the presence
22 and the absence of by-product benefits.

23

24 Biology of DSE fungi fits into the general definition of by-product mutualism and contrasts to key
25 characteristics of mycorrhizal fungi (in particular Glomeromycota, arbuscular mycorrhiza-forming
26 fungi) and, on the other hand, to free living saprotrophs in soil. The contrast to arbuscular mycorrhizal
27 symbiosis is of interest because of well-known co-colonization of arbuscular mycorrhizal fungi and
28 DSE fungi in herbaceous plants. DSE fungi possess intermediate characteristics which may be
29 indicative of differentiation from free saprotrophy towards mutualism in this fungal group
30 (Outstanding questions). Similar suggestions have been made also of other endophytic fungi, in
31 particular Sebaciniales [65,66]. Prescott et al. [48] recently hypothesised the role of plant excess
32 carbon as a driver of plant- soil interactions, especially in nutrient-limited conditions. The question
33 whether the amount of surplus carbon is enough to promote mycorrhizal fungi capable to degrade

1 these compounds remains to be solved. Excess carbon in plants, carbon in the litter and in particular
 2 exchanged carbon via mycorrhizal route give support to a hypothesis that excess carbon and by-
 3 products drive the evolution of mycorrhizal symbioses (Outstanding questions).

4

5 **Acknowledgements**

6 We thank Juhani Hopkins for checking the language of the manuscript and M.-A. Selosse and two
 7 anonymous referees for comments to the manuscript. This work was supported by the Academy of
 8 Finland, Grant Nos. 295976 and 326226 (to K.S.).

9 **References**

10

- 11 1 Jumpponen, A. and Trappe, J.M. (1998) Dark septate endophytes: a review of facultative biotrophic
 12 root-colonizing fungi. *New Phytol.* 140, 295–310
- 13 2 Jumpponen, A. et al. (2017) Biogeography of root-associated fungal endophytes. In *Biogeography*
 14 *of Mycorrhizal Symbioses* (Tedersoo, L. ed.), pp. 195-222, Springer
- 15 3 Newsham, K.K. (2011) A meta-analysis of plant responses to dark septate root endophytes. *New*
 16 *Phytol.* 190, 783-793
- 17 4 Mayerhofer, M.S. et al. (2013) The effects of fungal root endophytes on plant growth: a meta-
 18 analysis. *Mycorrhiza* 23, 119–128
- 19 5 Sieber, T.N and Grünig, C.R. (2013) Fungal root endophytes. In: *Plant Roots—The Hidden Half.*
 20 CRC Press, Taylor & Francis Group: Boca Raton, FL, USA, 38-1–38-49.
- 21 6 David, A.S. et al. (2016) Draft genome sequence of *Microdochium bolleyi*, a dark septate fungal
 22 endophyte of beach grass. *Genome Announc.* DOI: 10.1128/genomeA.00270-16
 23 (<https://mra.asm.org/>)
- 24 7 Schlegel, M. et al. (2016) Globally distributed root endophyte *Phialocephala subalpina* links
 25 pathogenic and saprophytic lifestyles. *BMC Genomics* 17, 1015
- 26 8 Knapp, D.G. et al. (2018) Comparative genomics provides insights into the lifestyle and reveals
 27 functional heterogeneity of dark septate endophytic fungi. *Sci.Rep.* 8, 6321
- 28 9 Clemmensen, K.E. et al. (2015) Carbon sequestration is related to mycorrhizal fungal community
 29 shifts during long-term succession in boreal forests. *New Phytol.* 205, 1525-1536
- 30 10 Sietiö, O.-M. et al. (2017) Ericoid plant species and *Pinus sylvestris* shape fungal communities in
 31 their roots and surrounding soil. *New Phytol.* 218, 738-751
- 32 11 Almario, J. et al. (2017) Root-associated fungal microbiota of nonmycorrhizal *Arabis alpina* and
 33 its contribution to plant phosphorus nutrition. *PNAS* 114, E9403-E9412
- 34 12 Yan D.F. et al. (2018) High-throughput eDNA monitoring of fungi to track functional recovery in
 35 ecological restoration. *Biol. Conserv.* 217,113-120
- 36 13 Li, X. et al. (2018) Dark septate endophytes isolated from a xerophyte plant promote the growth
 37 of *Ammopiptanthus mongolicus* under drought condition. *Sci. Rep.* 8, 7896
- 38 14 He, C. et al. (2020) Performance of enhancing licorice with dual inoculating dark septate
 39 endophytes and *Trichoderma viride* mediated via effects on root development. *BMC Plant Biol.*
 40 20, 325
- 41 15 Xu, R. et al. (2020) Evidence for a dark septate endophyte (*Exophiala pisciphila*, H93) enhancing
 42 phosphorus absorption by maize seedlings. *Plant Soil* 452, 249–266
- 43 16 Berthelot, C. et al. (2016) Plant growth promotion, metabolite production and metal tolerance of
 44 dark septate endophytes isolated from metal-polluted poplar phytomanagement sites. *FEMS*
 45 *Microbiol. Ecol.* 92: fiw144

- 1 17 Berthelot, C. et al. (2017) Differential growth promotion of poplar and birch inoculated with three
2 dark septate endophytes in two trace element-contaminated soils. *Int. J. Phytoremediation* 19,
3 1118-1125
- 4 18 Berthelot, C. et al. (2020) Unravelling the role of melanin in Cd and Zn tolerance and accumulation
5 of three dark septate endophytic species. *Microorganisms* 8, 537
- 6 19 Hou, L. et al. (2020) Dark septate endophytes improve the growth and the tolerance of *Medicago*
7 *sativa* and *Ammopiptanthus mongolicus* under cadmium stress. *Front. Microbiol.* 10: 3061
- 8 20 Yamaji, K. et al. (2016) Root fungal endophytes enhance heavy-metal stress tolerance of *Clethra*
9 *barbinervis* growing naturally at mining sites via growth enhancement, promotion of nutrient
10 uptake and decrease of heavy-metal concentration. *PLOS One* 11, e016089
- 11 21 Likar, M. (2018) Dark septate endophytic fungi and mycorrhizal fungi of trees affected by metal
12 pollution. In Pirttilä AM, Frank AC (eds) *Endophytes of forest trees*. Springer. p 119-137
- 13 22 Gonzalez Mateu et al. (2020) Dark septate endophyte improves salt tolerance of native and
14 invasive lineages of *Phragmites australis*. *ISME Journal* 14, 1943–1954
- 15 23 Yung, L., et al. (2021) Dark septate endophytes isolated from non-hyperaccumulator plants can
16 increase phytoextraction of Cd and Zn by the hyperaccumulator *Noccaea caerulescens*. *Environ.*
17 *Sci Pollut. Res.* DOI: 10.1007/s11356-020-11793-x (<https://link.springer.com>)
- 18 24 Mukasa Mugerwa, T.T. and McGee, P. (2016) Potential effect of melanised endophytic fungi on
19 levels of organic carbon within an Alfisol. *Soil research* 55: 245-252
- 20 25 Smith, S.E. and Read, D.J. (2008) *Mycorrhizal Symbiosis*. Academic press. 3rd edn.
- 21 26 Usuki, F. and Narisawa, K. (2007) A mutualistic symbiosis between a dark septate endophytic
22 fungus, *Heteroconium chaetospora*, and a nonmycorrhizal plant, Chinese cabbage. *Mycologia*
23 99, 175–184
- 24 27 Yakti, W. et al. (2019) Impact of dark septate endophytes on tomato growth and nutrient uptake.
25 *Plant Ecol. Divers.* 11, 637-648
- 26 28 Mandyam, K.G. et al. (2013) *Arabidopsis thaliana* model system reveals a continuum of responses
27 to root endophyte colonization. *Fungal Biol.* 117, 250–260
- 28 29 Mandyam, K.G. and Jumpponen A. (2014) Mutualism-parasitism paradigm synthesized from
29 results of root-endophyte models. *Front. Microbiol.* 5: 776
- 30 30 Caldwell, B.A. et al. (2000) Utilization of major detrital substrates by dark-septate root
31 endophytes. *Mycologia* 92, 230–232
- 32 31 Surono, S. and Narisawa, K. (2017) The dark septate endophytic fungus *Phialocephala fortinii* is
33 a potential decomposer of soil organic compounds and a promoter of *Asparagus officinalis*
34 growth. *Fungal Ecol.* 28, 1-10
- 35 32 Kiers, E.T. et al. (2011) Reciprocal rewards stabilize cooperation in the mycorrhizal symbiosis.
36 *Science* 333, 880–882
- 37 33 Noë, R. and Hammerstein, P. (1995) Biological markets. *Trends Ecol. Evol.* 10, 336–339
- 38 34 Noë, R. and Kiers, E.T. (2018) Mycorrhizal markets, firms and co-ops. *Trends Eco. Evol.* 33, 777-
39 789
- 40 35 Connor, R.C. (1995) The benefits of mutualism: a conceptual framework. *Biol. Rev.* 70, 427– 457
- 41 36 Leimar, O. and Hammerstein, P. (2010) Cooperation for direct fitness benefits. *Phil. Trans. R.*
42 *Soc. B* 365, 2619-2626
- 43 37 Bronstein, J. L. (ed) (2015) *Mutualism*. New York.
- 44 38 Harcombe, W.R. et al. (2018) Evolution of bidirectional costly mutualism from byproduct
45 consumption. *PNAS* 115, 12000-12004
- 46 39 Brown, J.L. (1983) Cooperation: a biologist's dilemma. *Adv. Study Behav.* 13, 1-37
- 47 40 Connor, R. C. (1986) Pseudo-reciprocity: investing in mutualism. *Anim. Behav.* 34, 1562-1584
- 48 41 Leimar, O. and Connor, R. (2003) By-product benefits, reciprocity and pseudo-reciprocity in
49 mutualism. In *Genetic & cultural evolution of cooperation*, Dahlem Workshop Report 90 (ed.
50 Hammerstein P.), pp. 203–222, Cambridge, MA: MIT Press

- 1 42 Saikkonen, K. et al. (2004) Evolution of endophyte-plant symbioses. *Trends Plant Sci.* 9, 275-280
- 2 43 Saikkonen, K. et al. (1998) Fungal endophytes: a continuum of interactions with host plants. *Annu.*
3 *Rev. Ecol. Syst.* 29,319–343
- 4 44 Johnson, N.C. (1997) Functioning of mycorrhizal symbiosis along mutualism-parasitism
5 continuum. *New Phytol.* 135, 575-586
- 6 45 Lynch, J.M. and Whipps, J.M. (1990) Substrate flow in the rhizosphere, *Plant Soil* 129: 1–10
- 7 46 Marschner, P. and Marschner, H. (2012) Mineral nutrition of higher plants. 3rd edn. Academic
8 Press.
- 9 47 Merino, C. et al., (2015) Soil carbon controlled by plant, microorganism and mineralogy
10 interactions. *J. Soil Sci. Plant Nutr.* 15, 321-332
- 11 48 Prescott, C.E. et al. (2020) Surplus carbon drives allocation and plant–soil interactions. *Trends*
12 *Ecol. Evol.* 35, 1110-1118
- 13 49 Knapp; D.G. and Kovács, G.M. (2016) Interspecific metabolic diversity of root-colonizing
14 endophytic fungi revealed by enzyme activity tests. *FEMS Microbiol. Ecol.* 92, fiw190
- 15 50 Challacombe, J.F. et al. (2019) Genomes and secretomes of Ascomycota fungi reveal diverse
16 functions in plant biomass decomposition and pathogenesis. *BMC Genomics* 20, 976
- 17 51 Jumpponen, A. et al. (1998) Mycorrhizal functioning of *Phialocephala fortinii* with *Pinus contorta*
18 on glacier forefront soil: interactions with soil nitrogen and organic matter. *Mycorrhiza* 7, 261-
19 265
- 20 52 Upson, R. et al. (2009) Nitrogen form influences the response of *Deschampsia antarctica* to dark
21 septate root endophytes. *Mycorrhiza* 20, 1–11
- 22 53 Postma, J. et al. (2007) Root colonisation by arbuscular mycorrhizal, fine endophytic and dark
23 septate fungi across a pH gradient in acid beech forests. *Soil Biol. Biochem.* 39, 400–408
- 24 54 Francini, G. et al. (2014) Arbuscular mycorrhizal fungal community divergence within a common
25 host plant in two different soils in a subarctic Aeolian sand area. *Mycorrhiza* 24, 539–550
- 26 55 Kauppinen, M. et al., (2014) Contrasting preferences of arbuscular mycorrhizal and dark septate
27 fungi colonizing boreal and subarctic *Avenella flexuosa*. *Mycorrhiza* 24, 171-177
- 28 56 Huusko, K. et al. (2017) A shift from arbuscular mycorrhizal to dark septate endophytic
29 colonization in *Deschampsia flexuosa* roots occurs along primary successional gradient. .
30 *Mycorrhiza* 27, 129-138
- 31 57 Hill, P.W. et al. 2019 Angiosperm symbioses with non-mycorrhizal fungal partners enhance N
32 acquisition from ancient organic matter in a warming maritime Antarctic. *Ecol. Lett.* 22: 2111–
33 2119
- 34 58 Hadacek, F. and Kraus, G.F. (2002) Plant root carbohydrates affect growth behaviour of
35 endophytic microfungi. *FEMS Microbiol. Ecol.* 41, 161–170
- 36 59 Martino, E. et al. (2018) Comparative genomics and transcriptomics depict ericoid mycorrhizal
37 fungi as versatile saprotrophs and plant mutualists. *New Phytol.* 217, 1213-1229
- 38 60 Wu, F. L et al. (2020) Novel dark septate fungal endophyte positively affected blueberry growth
39 and changed the expression of plant genes involved in phytohormone and flavonoid biosynthesis.
40 *Tree Physiol.* 40, 1080-1094
- 41 61 Zhang, Y. et al. (2009) Multi-locus phylogeny of Pleosporales: a taxonomic, ecological and
42 evolutionary re-evaluation. *Stud. Mycol.* 64: 85-102S5.
- 43 62 Schoch, C. L. et al. (2009) The Ascomycota tree of life: a phylum-wide phylogeny clarifies the
44 origin and evolution of fundamental reproductive and ecological traits. *Syst. Biol.* 58, 224-239
- 45 63 Maynard Smith, J. and Szathmáry, E. (1995) The major transitions in evolution. Oxford University
46 Press.
- 47 64 Hoeksema, J.D. and Bruna, E.M. (2015) Context-dependent outcomes of mutualistic interaction.
48 In *Mutualism* Bronstein J.L. (ed) pp. 181- 202. Oxford.
- 49 65 Selosse, M.-A. et al. (2009) Do Sebaciales commonly associate with plant roots as endophytes?
50 *Mycol. Res.* 113, 1062-1069

- 1 66 Selosse, M.-A. et al. (2018) Time to re-think fungal ecology? Fungal ecological niches are often
2 prejudged. *New Phytol.* 217, 968–972
- 3 67 Hoeksema, J.D. et al. (2018) Evolutionary history of plant hosts and fungal symbionts predicts the
4 strength of mycorrhizal mutualism. *Commun. Biol.* 1, 116
- 5 68 Chamberlain, S.A. et al. (2014) How context dependent are species interactions? *Ecol. Lett.* 17,
6 881–890
- 7 69 Chomicki, G. et al. (2020) The evolution of mutualistic dependence. *Annu. Rev. Ecol. Evol. Syst.*
8 51, 409–432
- 9 70 Hoystedt, G.A. et al. (2018) A mycorrhizal revolution. *Curr. Opin. Plant Biol.* 44, 1-6
10

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25

Figure legends

Figure 1. Dark septate endophyte (DSE) hyphae in the root cortex of *Deschampsia flexuosa* Root preparation bleached and stained with trypan blue. 400x magnification in compound (light) microscope.

Figure 2. Microsclerotium in the root cortex cell of *Deschampsia flexuosa* Root preparation bleached and stained with trypan blue. 400x magnification in compound (light) microscope.

Figure 3. By-product mutualism in DSE symbiosis Deposits from the host plant (C) are utilized by dark septate endophytic (DSE) fungi in soil. Activity of DSE fungi associated to host plant roots increases nutritional and potentially other benefits (N) for the plant. This figure was partly created using BioRender (<https://biorender.com/>).

Figure 4. Symbiont derived benefit for the plant in relation to the plant investment A schematic presentation on symbiont-derived benefit for the plant (solid curves) in relation to the plant investment for maintenance of the symbiosis (dashed line). Open dots show the worst and closed dots the best of the plant benefit-cost balance for the plant profits. Investment optima, “best solutions”, for the plant: black dot = no investment, red dot = highest investment level, green dot = investment level with some by-product benefits on the symbionts (if a minimum occurs at lower investment level but above 0). Blue dots = by-product benefits received without any investments from the plant partner. This figure was partly created using BioRender (<https://biorender.com/>).

1 **Table 1.** Contrasting symbiosis-related characteristics in plant-fungal associations with mycorrhizal
 2 fungi, dark septate endophytes (DSE) and free saprotrophs

Symbiosis-related characteristics	Mycorrhiza ^a	DSE	Free saprotrophs
Potential role of by-products in association	Low	High	High
Structural investment cost	High	Low	No or low
Symbiont derived benefit	High ^b	Intermediate	No or low
Dependency on host	High-intermediate	Low	Low
Host-Specificity	High ^b	Intermediate	Low
Taxonomic groups involved	Ascomycota, Basidiomycota, Glomeromycota, Mucoromycotina ^c	Ascomycota	Diverse, with potentially all phyla represented

3 ^aEricoid, orchid and sebacinoid mycorrhizal fungi also have advanced saprotrophic capacity.

4 ^bBenefit and specificity in mycorrhizal symbioses varies according to the mycorrhizal type.

5 ^cHoystedt et al. [70]

6

7

8

9

10

11

1 **Box 1. Dark septate endophytic (DSE) fungi**

2 DSE fungi colonize living plant roots by melanized and hyaline septate hyphae (septa = cell wall,
3 hyaline = colourless) (Figure 1). In addition, microsclerotia, intraradical resting structures of the
4 fungus are characteristic to DSE (Figure 2). Based on root colonization morphology alone, DSE
5 cannot be identified, therefore investigations by laboratory and molecular techniques are needed [5].
6 But distinctive hyphal morphology inside young, healthy roots without visible symptoms in host
7 plants is considered indicative of DSE colonization.

8 Functioning of the DSE symbiosis in the roots has remained obscure, i.e. whether or not fungal-
9 mediated exchange of carbon and nutrients takes place between the host and the fungi. Specialized
10 structures for carbon and nutrient exchange between the plant and the fungus, which are typical to
11 mycorrhizal symbiosis, do not exist in DSE.

12 DSE fungi have highly developed capacity to degrade organic matter (saprotrophy).

13 DSE fungal cultures can produce conidia (= asexual spores) and certain macrofungi, such as
14 *Mollisia* and *Pyrenopeziza*, have an association to DSE. However, there is no comprehensive
15 knowledge of life cycles of DSE fungi in the wild.

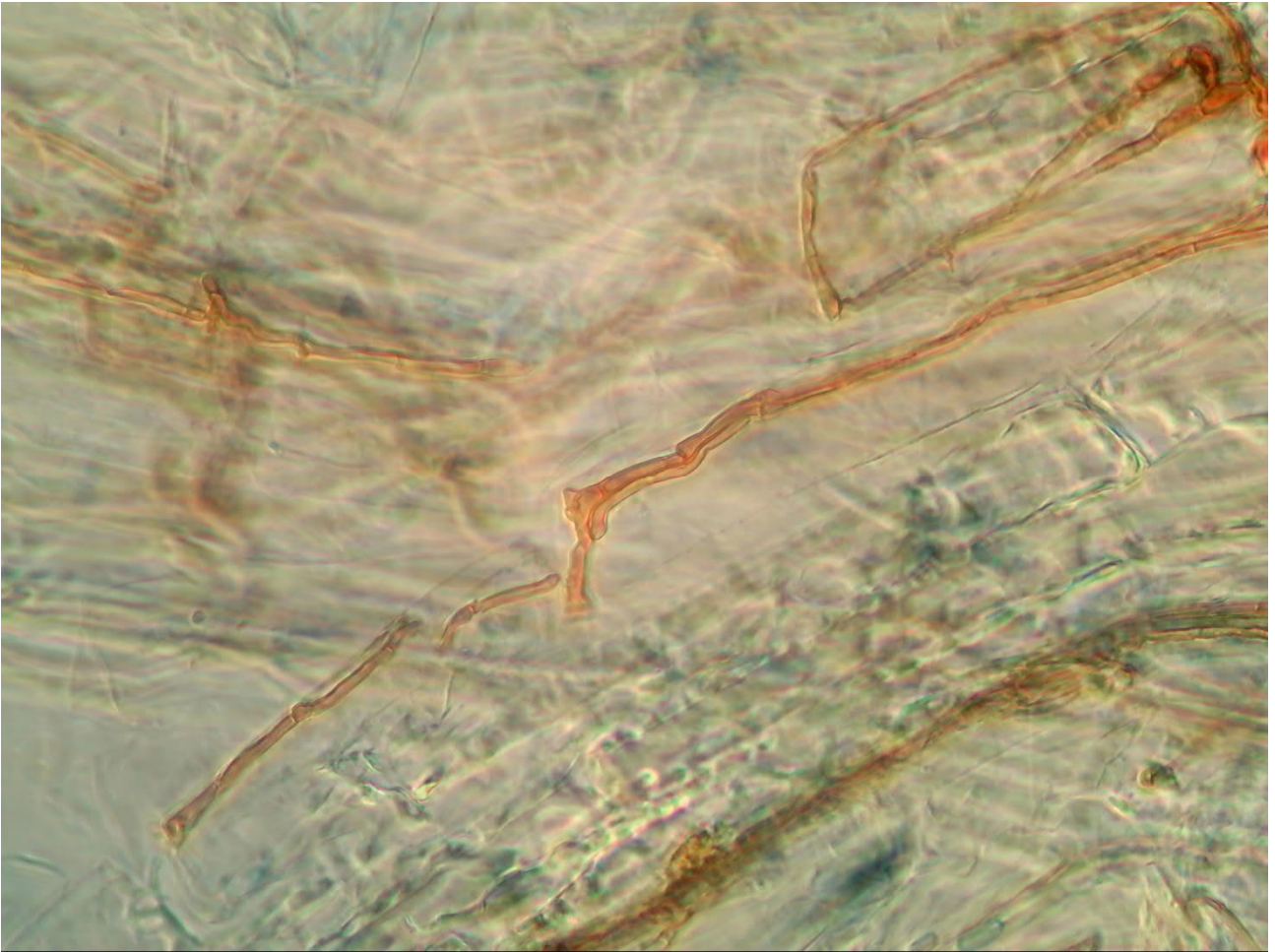
16

17

18

1 Figure 1.

2



3

4

5

1 Figure 2.

2



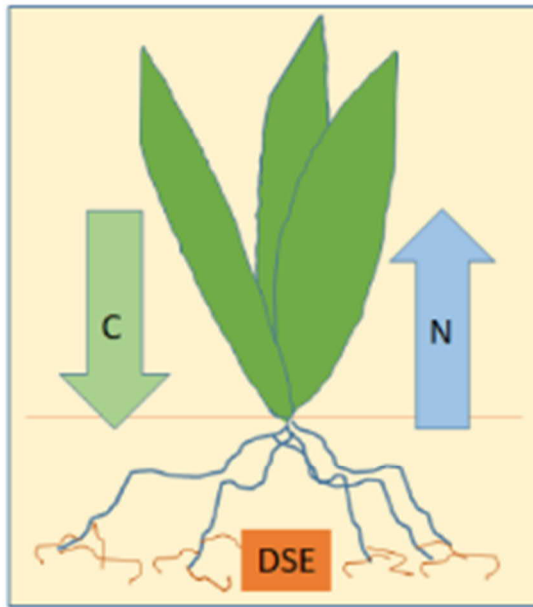
3

4

5

1 Figure 3.

2



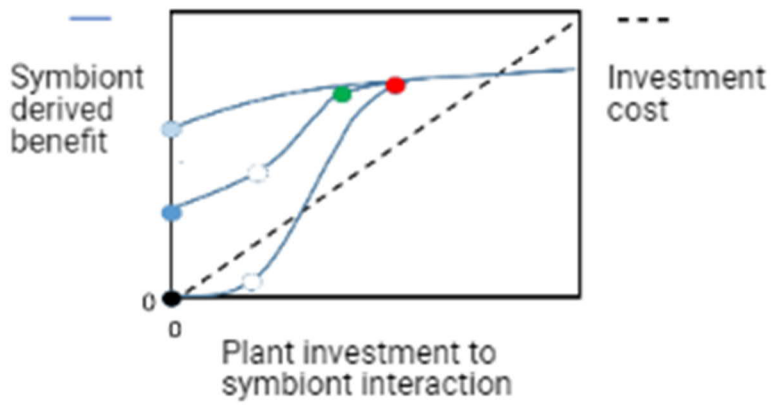
3

Created in BioRender.com 

4

1 Figure 4.

2



3

Created in BioRender.com bio

4

5