

1 Restraining fluoride loss from  $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$   
2 upconverting nanoparticles in aqueous environments  
3 using crosslinked poly(acrylic acid)/poly(allylamine  
4 hydrochloride) multilayers

5

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16 **KEYWORDS:** layer-by-layer; polyelectrolytes; crosslinking; nanoparticles; upconversion  
17 luminescence

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21 **ABSTRACT**

22 The use of upconverting nanoparticles in various applications in aqueous media relies on their  
23 surface modifications as most synthesis routes yield hydrophobic particles. However, introducing  
24 upconverting nanoparticles in aqueous solutions commonly results in the quenching of their  
25 luminescence intensity and in the worst case, disintegration of the nanoparticles. We demonstrate  
26 the use of poly(acrylic acid) and poly(allylamine hydrochloride) as a protecting layer-by-layer  
27 coating for the upconverting NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> nanoparticles. The formation and crosslinking of  
28 the bilayer coating was confirmed with Fourier transform infrared spectroscopy, thermal analysis  
29 and zeta potential. The release of internal fluoride ions from the nanoparticle structure and  
30 subsequent particle disintegration was decelerated especially by crosslinking the bilayer coating  
31 on the surface. In addition, we studied the effect of the coating on the upconversion luminescence  
32 properties and learned that with additional fluoride ions present during the layer-by-layer assembly  
33 the most intense enhancement in the luminescent intensity is obtained. This is due both to not  
34 allowing the disintegration of the particles during the surface modification process as well as  
35 preventing the water molecules accessing the surface by crosslinking the bilayer coating.

36

37 **INTRODUCTION**

38 The increasing research on the use of upconversion nanomaterials in various applications such as  
39 biosensors [1], biomedical assays and imaging [2,3] and in theranostics [4] is based on the  
40 materials' unique property of converting low energy radiation (mainly near infra-red, NIR) into  
41 higher energy radiation (visible light) [5,6]. The NIR excitation needed for upconversion  
42 nanomaterials provokes less photodamage in the imaged tissue in comparison with the  
43 conventional ultraviolet (UV) excitation. Furthermore, the lack of autofluorescence in the

44 background benefits especially the biomedical field enabling faster processes due to not needing  
45 time-resolved measurements [7]. While the development of smaller nanoparticles has widened  
46 their use range in research [8,9] there are drawbacks that need addressing. Hexagonal  $\beta$ -  
47  $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$  is probably the most widely used upconverting nanoparticle (UCNP) nowadays  
48 due to its superiority as the most efficient upconversion material [10,11]. However, one of the  
49 most crucial drawbacks in the  $\text{NaYF}_4$  materials is the quenching of upconversion luminescence in  
50 aqueous environments through energy migration within the sensitizing ytterbium ions in the  
51  $\text{NaYF}_4$  lattice and the large surface area that can be full of impurities [12–14]. In addition to energy  
52 migration, recent studies suggest that this luminescence quenching is also caused by disintegration  
53 of the  $\text{NaYF}_4$  UCNPs in aqueous media [15,16].

54 Methods of enhancing the performance of  $\text{NaYF}_4$  UCNPs have been developed, such as  
55 manufacturing various types of core-shell materials [17,18] and using metal frameworks near the  
56 particle to induce plasmonic effects [19,20]. However, as long as the fluoride ions remain on the  
57 surface of the nanoparticle its disintegration is probable when the nanoparticles are in direct contact  
58 with water. Also, because most of the common synthesis methods produce hydrophobic  
59 nanoparticles [7,21], multiple surface functionalization steps need to be taken to enable the  
60 dispersion of the particles into water [2,22]. A few surface modification strategies for hindering  
61 the disintegration of UCNPs have already been published but none have been able to prevent the  
62 disintegration completely [23,24].

63 In the current study, we investigated the surface functionalization of UCNPs using a layer-by-layer  
64 method to produce bilayers with self-assembled negatively and positively charged  
65 polyelectrolytes, namely poly(acrylic acid) (PAA) and poly(allylamine hydrochloride) (PAH)  
66 [25,26]. Nanoparticles have been used as a template for the layer-by-layer method previously when

67 various types of bilayers have been manufactured [27–29]. However, the use of the layer-by-layer  
68 method in the surface functionalization of UCNPs has not been studied in detail as the bilayer  
69 formation is affected by a multitude of conditions during the process. Previously, it has been  
70 observed that by employing crosslinked PAA/PAH layers with a  $\text{Cu}^{2+}$ -template on a flat alumina  
71 surface it was possible to reduce the anion flux through the alumina membrane [30]. We wanted  
72 to investigate if cross-linking could be used to reduce or even prevent the outward fluoride flux  
73 from the  $\text{NaYF}_4$  particles that would cause the disintegration of UCNPs in aqueous environments.  
74 The bilayer coating formation using PAA and PAH was studied with selected ionic concentrations  
75 at pH 5.5 which provides enough charge for both polyelectrolytes. Coating was also manufactured  
76 in the presence of an additional fluoride source to investigate the disintegration during the layer-  
77 by-layer coating cycles. Fourier transform infrared (FT-IR) spectroscopy, thermal analysis and  
78 zeta potential measurements were used to confirm the formed coating and the crosslinking of the  
79 bilayer structure. To see how the bilayers and their crosslinking affected the disintegration of the  
80 nanoparticles in aqueous environments the concentration of released internal fluoride was  
81 measured. Changes in the upconversion luminescence properties of the core materials was studied  
82 using a 973 nm excitation suitable for the excitation of  $\text{Yb}^{3+}$  and  $\text{Er}^{3+}$  ions.

83

## 84 MATERIALS AND METHODS

### 85 Reagents.

86 Poly(acrylic acid) (PAA;  $M_w \approx 100,000$ ),  $(\text{C}_3\text{H}_4\text{O}_2)_n$ , Aldrich), poly(allylamine hydrochloride)  
87 (PAH;  $M_w \approx 50,000$ ,  $[\text{CH}_2\text{CH}(\text{CH}_2\text{NH}_2 \cdot \text{HCl})_n$ , Aldrich), sodium chloride (NaCl 99.5 %, J.T.  
88 Baker), sodium fluoride (NaF, >99 % Fluka), sodium nitrate ( $\text{NaNO}_3$ , 99.5 % Riedel-de Haen).  
89 Absolute ethanol (>99.5 %, Altia) was used as received.

90

91 **Materials preparation.**

92 The  $\beta$ -NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> ( $x_{\text{Yb}}$ : 0.17,  $x_{\text{Er}}$ : 0.03) nanoparticles (size *ca.* 19\*23 nm) used as the core  
93 material were prepared with the synthesis procedure reported previously [31]. The oleic acid  
94 present at the nanoparticle surface was removed with a previously described acidic treatment [32].  
95 The coating solutions were 10 mM of used polyelectrolyte (in reference to the monomer  
96 concentration of the polyelectrolyte) solubilized in 0.1 or 0.2 M NaCl (aq). The pH of the solutions  
97 was 5.5. An additional layer-by-layer assembly using ionic concentration of 0.1 M NaCl was made  
98 with extra fluoride present. In this case an additional 10 mM of NaF was added into each used  
99 solution including the water used for washing.

100 The coating cycle of the nanoparticles was the same as reported previously [29]. The cycle  
101 involved dispersing the core nanoparticles into the desired coating solution and ultrasonicing for  
102 two minutes and washing with quartz distilled water twice to prepare half a bilayer. One, three and  
103 five bilayers were manufactured to ensure layer formation. Also an additional PAA layer was  
104 added on top with a similar procedure to provide attaching sites for biomolecules and to ensure  
105 suspension in water based solutions. This makes the total number of bilayers 1.5, 3.5 and 5.5. The  
106 crosslinking of the formed bilayers was performed at 180 °C (ramp step 2 °C/min) for two hours  
107 in a nitrogen atmosphere. The outermost layer of PAA was expected not to take part in the  
108 crosslinking process to a great extent thus leaving negative carboxyl ends as attaching sites to the  
109 surface.

110

111 **Characterization**

112 The core particles' crystal structure was determined at room temperature with X-ray powder  
113 diffraction (XRD) using a Huber G670 image plate Guinier camera (Cu  $K_{\alpha 1}$  radiation, 1.5406 Å)  
114 with a  $2\theta$  range of 4-100° (step 0.005°). Data collection time was 30 min followed by 10 data  
115 reading scans of the image plate. From this data the crystallite size of the core material was  
116 calculated with the Scherrer formula using reflections (002) and (200) for the thickness and width  
117 of the hexagonal faces, respectively.

118 The presence of the polyelectrolytes was studied with FT-IR spectra using Bruker Vertex 70 MVP  
119 Star Diamond setup with 256 scans between 450 and 4500  $\text{cm}^{-1}$ . The resolution was 4  $\text{cm}^{-1}$ . The  
120 thermal behavior of the coated materials was studied with thermogravimetric analysis (TGA) and  
121 differential scanning calorimetry (DSC) using one measurement per sample with a TA Instruments  
122 SDT Q600 TGA-DSC apparatus. The measurements were made between 35 and 600 °C with the  
123 heating rate of 10 °C/min using flowing air sphere (100 ml/min). The error for weight is *ca.* 1 %  
124 in this setup. The Zeta potential of the materials was measured from three parallel measurements  
125 with Malvern Zetasizer Nano-ZS equipment. The concentration of aqueous solutions was 100  
126  $\mu\text{g/ml}$  with pH of *ca.* 6. The images of the coated nanoparticles were obtained with JEM-1400 Plus  
127 transmission electron microscopy (TEM) using OSIS Quemesa 11 Mpix bottom mounted digital  
128 camera. The nanoparticles were suspended into ethanol and dried on a lacey carbon grid and then  
129 imaged with acceleration of 60 kV.

130 Disintegration of the nanoparticles was studied using fluoride selective electrode was studied with  
131 one measurement per sample. Known amount (*ca.* 2 mg) of the nanoparticles was combined with  
132 1 ml of water and sealed in a dialysis tube (Orange Scientific, Regenerated cellulose dialysis tubing  
133 MWCO 3500). The dialysis tube with the contents was then immersed in aqueous solution of 0.1  
134 M  $\text{NaNO}_3$  (total volume 50 ml) with the ion selective and reference electrodes. The data was

135 collected automatically once every minute during the 24 hour measurement window. When the  
136 measurements were made with external  $10^{-5}$  M NaF it was added into all solutions including the  
137 one inside the dialysis tube.

138 The upconversion luminescence spectra were measured at room temperature with an Avantes  
139 Avaspec HS-TEC spectrometer. A fiber-coupled continuous NIR laser diode IFC-975-008-F  
140 (Optical Fiber Systems) with the excitation wavelength of 973 nm ( $10\,270\text{ cm}^{-1}$ ) was used as an  
141 excitation source with 5000 mA corresponding  $9.1\text{ Wcm}^{-2}$ . Dry nanomaterials were held inside a  
142 rotating capillary tube and measured with 20 averaging scans, 20 ms each. After the sample a  
143 short-pass filter with a cutoff of 750 nm (Newport) was used to exclude excitation radiation. The  
144 emission was collected at a  $90^\circ$  angle to the excitation and directed to the spectrometer with an  
145 optical fiber.

146

## 147 **RESULTS AND DISCUSSION**

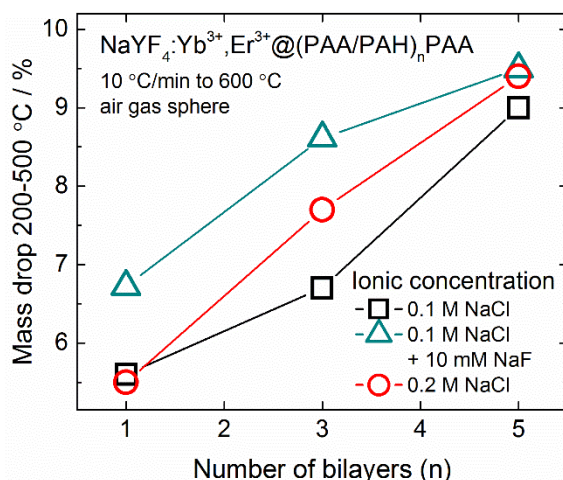
### 148 **Layer-by-layer assembly on the nanoparticles**

149 The  $\beta\text{-NaYF}_4\text{:Yb}^{3+},\text{Er}^{3+}$  upconversion nanomaterials used as core materials in the layering process  
150 were of hexagonal form and *ca.*  $19*23\text{ nm}$  in size (Table S1). The removal of oleic acid surface  
151 was confirmed with the FT-IR measurements as the signals of the asymmetric and symmetric  
152 vibrations of  $\text{COO}^-$  could be seen to decrease (Figure S1).

153 When the FT-IR spectra of the coated nanoparticles were measured it was observed that with all  
154 materials the characteristic vibrations from PAA and PAH electrolytes were present in the  
155 materials (Figure S2) [33,34]. This could be observed from the presence of asymmetric and  
156 symmetric vibrations from carboxyl groups  $\text{COO}^-$  (*ca.*  $1400$  and  $1550\text{ cm}^{-1}$ ) and  $\text{COOH}$  ( $1710\text{ cm}^{-1}$ )  
157  $^1$ ) from PAA and a broader band from  $\text{NH}_2$  bending in PAH at  $800\text{-}900\text{ cm}^{-1}$ . The formation is

158 considered to be similar to the bilayer formation between PAA and PAH established on a planar  
159 surface [30,33,35,36].

160



161

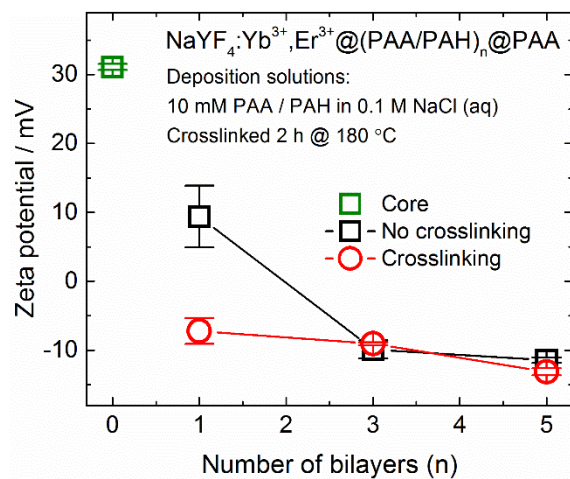
162 Figure 1. The mass drop of the coated UCNPs between 200 and 500 °C calculated from the thermal  
163 analysis data.

164

165 The thermal analysis of the coated nanoparticles suggested that the formation of bilayers varied  
166 between the deposition conditions (Figure 1, S3). It seemed that it is the most linear with the ionic  
167 concentration of 0.2 M. The percentage of the removed mass from the coated materials made with  
168 additional 10mM fluoride was higher from those prepared without fluoride. This suggested that in  
169 addition to interlocked water there were also residual fluoride ions from the additional fluoride  
170 solution in the bilayer structure that evaporates at higher temperatures behaving similarly to  
171 amorphous NaF that is present in the annealing of cubic NaYF<sub>4</sub> structure [37]. However, the mass  
172 drop with five bilayers was similar in all of the coated materials and the difference in the behavior  
173 with the low number of bilayers is expected to arise only from the differences in the layer formation  
174 process. It could be that the higher ionic concentration is more efficient in stacking the  
175 polyelectrolyte to more compact and dense structures and thus making them easier to cover the



176 nanoparticle surface without additional branched structures that might hinder uniform layer  
177 production and cause diversity into layers [38].

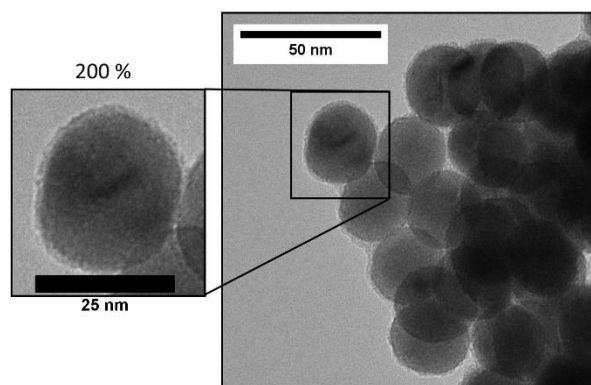


178  
179 Figure 2. Zeta potential of the as-prepared and crosslinked coated nanomaterials prepared with  
180 ionic concentration of 0.1 M NaCl.

181  
182 When zeta potential was considered, the nanomaterials regardless of the ionic concentrations  
183 during the layering process showed a similar trend in the surface behavior. The zeta potential  
184 changed from the core materials' *ca.* 30 mV to the *ca.* -10 mV of the materials having five bilayers  
185 (Figure 2, S4). Also the zeta potential of the materials deposited with additional 10 mM fluoride  
186 are similar to those deposited at the same conditions without the fluoride. This suggests that the  
187 amount of extra fluoride is low enough not to affect the bilayer formation. With all of the coated  
188 nanomaterials the first bilayer showed the highest variation during measurement. This could  
189 indicate that the first bilayer structure is not as uniform as the following layers.

190  
191 The TEM imaging of the coated nanoparticles was used to study if we observe the bilayer  
192 formation on the surface (Figure 3). As both polyelectrolytes used in the coating contain only very  
193 light elements in comparison with those in the NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> core particles the imaging was

194 made using a lacey carbon grid with low voltage (60 kV) to minimize the background and to give  
195 more contrast to the surface. Unfortunately the use of a carbon grid as a template aggregates the  
196 nanoparticles on the grid boundaries similarly as observed on the previous layer-by-layer coated  
197 nanoparticles [24,29]. While the exact amount of the bilayers cannot be seen from the TEM  
198 images, the coating was visible when the material coated with 5.5 bilayers of (PAA/PAH) was  
199 imaged. The thickness of the coating is *ca.* 2 nm which is similar to those obtained previously [29].

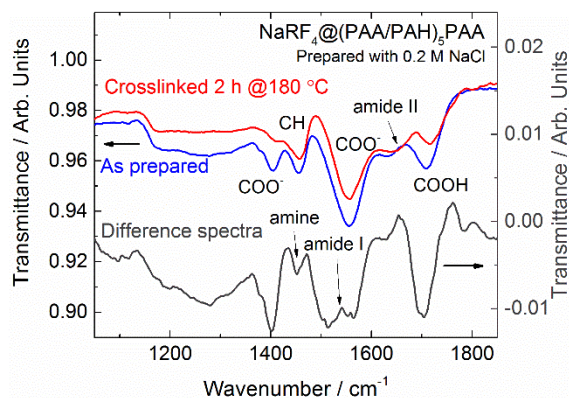


200  
201 Figure 3. TEM image of the coated nanoparticles with 5.5 bilayers of (PAA/PAH) on the surface.  
202 Scale bar is 50 nm and the inset zoom 200 % with a 25 nm scale bar.

203  
204 **Crosslinking of the bilayers**  
205 The thermal crosslinking between the carboxyl group of PAA and amine group of PAH has been  
206 successfully demonstrated in the literature between 130-215 °C [30,33,35,36]. Due to low amount  
207 of sample material we chose to test the crosslinking by heating the nanomaterial (*ca.* 50 mg) for  
208 two hours at both 180 and 200 °C in N<sub>2</sub> atmosphere (Figure S5). This was done with material  
209 coated with five bilayers of PAA/PAH without the additional PAA on the surface to remove the  
210 effect of free carboxyl acid (-COOH) in the FT-IR spectra. From the FT-IR spectra it could be  
211 observed that crosslinking occurred at both temperatures but no clear difference between these two

212 temperatures could be found. The damage to the surface modifications is expected due to the  
213 thermal analysis of the coated materials in which the mass drop is seen to increase already at the  
214 200 °C. As the upconversion luminescence of the material crosslinked at 200 °C was weaker than  
215 that of 180 °C material (Figure S5) the latter was chosen as crosslinking temperature without  
216 further studies.

217



218

219 Figure 4. FT-IR spectra of the as-prepared and crosslinked coated nanomaterials prepared with  
220 ionic concentration of 0.2 M NaCl and their difference spectra.

221

222 When a difference FT-IR spectra is drawn from the 5.5 bilayer coated as-prepared and crosslinked  
223 nanomaterials it can be observed that the carboxyl and amine related vibrations (1400, 1570, 1710  
224 and 800  $\text{cm}^{-1}$ , respectively) are reduced and the amide related vibrations at *ca.* 1550 and 1660  $\text{cm}^{-1}$   
225 are increasing suggesting that the crosslinking was successful (Figure 4) [30,33]. From the FT-  
226 IR spectra of the crosslinked nanomaterials it could be seen that there are still remaining  $\text{COO}^-$   
227 vibrations from the final PAA layer on the surface as expected (Figure S6).

228 After crosslinking, a change could be observed in the zeta potential at the first bilayer of the  
229 materials as it drops *ca.* 15 mV with both ionic concentrations in all materials (Figure 2, S4). This  
230 suggested that the crosslinking of the first bilayer removes the positive charge inside the

231 polyelectrolyte multilayers. With the higher number of bilayers the change in zeta potential is not  
232 as big suggesting that even without the crosslinking the surface structure is more defined with the  
233 higher number of bilayers.

234

### 235 **Particle disintegration**

236 The particle disintegration was monitored in aqueous environments using a fluoride selective  
237 electrode. We also studied the disintegration behavior in the presence of a low concentration of  
238 fluoride and in those measurements an external  $10^{-5}$  M of NaF was introduced into the aqueous  
239 solutions. The external fluoride concentration is expected to decelerate the disintegration process.  
240 Lahtinen et al. have previously studied the fluoride concentration effect on the disintegration of  
241 the nanoparticles up to 100 mM [15]. However, this concentration is already too high for the  
242 fluoride electrode response and thus unavailable for our measurement. The external fluoride was  
243 then subtracted from the data to obtain the actual release of the internal fluoride from the  
244 nanoparticles. From this data, a 1<sup>st</sup> order exponential fit was made to determine the total  
245 disintegration of the particle as the disintegration is expected to slow down as the equilibrium is  
246 reached with the fluoride concentration of the solution. However, it must be noted that in the case  
247 of the coated nanoparticles this is expected to be more complicated because the concentration of  
248 the fluoride ions is expected to vary when different parts of the environments are considered (*e.g.*  
249 direct particle surface, the bilayer structure and the whole aqueous solution). For the fits the first  
250 30 minutes were discarded since it takes 30 minutes for the instantly removed fluoride ions to  
251 reach the electrode through the dialysis barrier. The internal fluoride release rate during the  
252 measurement was estimated from the exponential fit derivative.

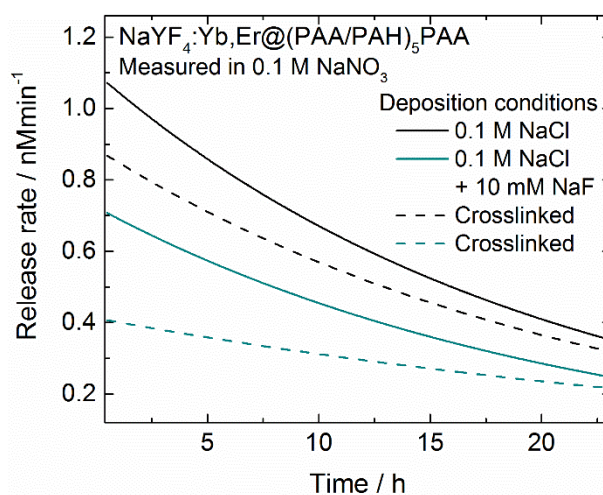
253

254 The used core materials released between 1.7-2.3 mol-% of fluoride ions during the 24 h  
255 measurement window. The differences in the disintegration are expected to arise from the size of  
256 the particles (the largest particle disintegrated the least) and from the possible differences in the  
257 remaining surface impurities within the core materials. The total extrapolated disintegration was  
258 from 2.7 to 3.4 mol-% for the core materials (Table S2). Using external fluoride the disintegration  
259 was decelerated slightly (*ca.* 1 mol-%). The behavior of the core particle is similar to the data  
260 obtained by Lisjak et al. where in the course of three days the fraction of fluoride dissolved in  
261 water was 3.7 mol-% [39]. However, they used higher concentration of UCNPs (1 mg/ml) which  
262 is expected to make the UCNPs disintegrate less than those made with lower UCNP concentration  
263 [15].

264  
265 From the coated nanomaterials the 5.5 bilayers of PAA/PAH were the most successful in  
266 decelerating the fluoride release from the nanoparticle surface (Figure S7). In addition, already the  
267 first 1.5 bilayers were also successful but not as efficient. The biggest differences in the internal  
268 fluoride release were obtained from the higher number of coated bilayers. Crosslinking of the  
269 bilayers decelerated the disintegration even more during the measurement window, but in some  
270 cases the overall extrapolated disintegration was similar to that of the as-prepared materials  
271 regardless of the crosslinking. The most efficient coatings in decelerating the disintegration were  
272 5.5 bilayers prepared with ionic concentration of 0.2 M NaCl with or without the crosslinking.

273  
274 The fluoride release rate curves showed that for these efficient coatings the release of the internal  
275 fluoride ions is decelerated faster than for the others (Figure 5, S8). However, the curves of the  
276 coated materials prepared with 0.1 M NaCl using additional 10 mM of NaF during the coating

277 suggested that the release of the internal fluoride ions was the slowest with their crosslinked 5.5  
278 bilayer material even though the overall release during the measurement window is slightly higher.  
279 This suggests that during the coating with additional fluoride some of the fluorides were  
280 accommodated by the positive amine sites of PAH creating a higher fluoride surface concentration  
281 within the formed bilayer structure and thus decelerating the release of the internal fluorides from  
282 the nanoparticle. Especially during the first five hours the difference between the fluoride release  
283 rates between the crosslinked and unlinked materials was significant. This suggests that the  
284 crosslinking can benefit specially the applications with short measurement windows.



285  
286 Figure 5. Fluoride release rate from the 5.5 bilayer coated nanomaterials made with 0.1 M NaCl  
287 measured in 0.1 M NaNO<sub>3</sub>.

288  
289 Overall, the disintegration was slower from the coated nanomaterials prepared with additional  
290 fluoride present. We believe that when UCNPs are coated while there is additional fluoride present  
291 the rapid first hydrolysis of the fluorides on the particle surface [39] can be completely prevented  
292 during the coating process. This means that the disintegration has to start from an intact UCNP  
293 surface during the fluoride release measurement. As the surface is intact the process is slower. This

294 means that without additional fluoride present during the coating the surface ions are already lost  
295 during the modification steps resulting in weakened surface properties.

296 The equilibrium state of the fluoride concentration in  $\text{NaNO}_3(\text{aq})$  is reached fastest (*ca.* 75 hours)  
297 with the materials coated with 5.5 bilayers of (PAA/PAH) using 0.2 M NaCl concentration  
298 regardless of the crosslinking (Figure S9). With the used cores and materials coated with 5.5  
299 bilayers using the 0.1 M NaCl concentration the disintegration continued past the 75 hours and the  
300 equilibrium was reached only after 100 hours. The crosslinked materials had lower rate constants  
301 than did the non-crosslinked ones (Table S3) confirming that the internal fluoride release in them  
302 is slower than in those without crosslinking. However, no other trends could be observed with the  
303 coated materials.

304  
305 Table 1. Disintegration of UCNPs coated with 5.5 bilayers with selected deposition conditions  
306 measured with 0.1 M  $\text{NaNO}_3(\text{aq})$  with or without external  $10^{-5}$  M  $\text{NaF}(\text{aq})$  at 23 hours and in  
307 extrapolated equilibrium (200 h).

Deposition conditions with 5.5 bilayers	x (F) @ 23 h / mol-%		x(F) @ 200 h / mol-%	
	no fluoride	external fluoride	no fluoride	external fluoride
0.1 M NaCl	1.7	1.1	2.4	1.7
above crosslinked	1.4	0.5	2.2	1.6
0.1 M NaCl + 10mM NaF	1.3	1.5	1.9	1.9
above crosslinked	1.1	0.8	2.1	1.9
0.2 M NaCl	1.4	1.0	1.7	1.4
above crosslinked	1.3	0.8	1.8	1.3

308  
309 When measurements from the materials having 5.5 bilayers on the nanoparticle surface were  
310 conducted in the presence of external  $10^{-5}$  M NaF in the nitrate solution it was observed that the  
311 internal fluoride release with the coated materials decreased (Table 1, Table S2, Figure S7). This

312 is in agreement with the previously reported studies in which presence of 1 mM KF during the  
313 measurement prevented the particle disintegration [15]. However, it was interesting that with  
314 external fluoride present at the measurement with the nanomaterials coated with additional fluoride  
315 the release of the internal fluoride increased. It may be that this is due to such fluoride ions that  
316 have been captured within the coating layers during the coating process. Their release is seen as  
317 an apparent increase in the fluoride disintegration. This is expected to happen also with  $\text{NaNO}_3(\text{aq})$   
318 and thus the observed released  $x(\text{F})$  for the as prepared materials made with the presence of  
319 external fluoride may be misleading. However, during the crosslinking some of this additional  
320 fluoride in the structure is expected to evaporate [37].

321

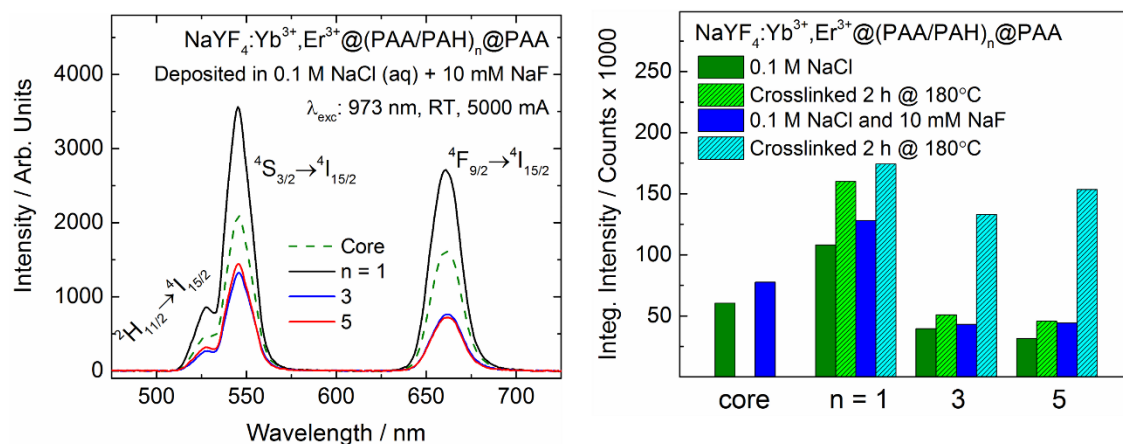
### 322 **Upconversion luminescence properties**

323 All of the materials produced the desired visible upconversion luminescence from  $\text{Er}^{3+}$  ions in  
324 green ( ${}^2\text{H}_{11/2}, {}^4\text{S}_{3/2} \rightarrow {}^4\text{I}_{15/2}$ ) and red ( ${}^4\text{F}_{9/2} \rightarrow {}^4\text{I}_{15/2}$ ) when excited at 973 nm ( $10\,280\text{ cm}^{-1}$ ). This is  
325 made possible through the energy transfer upconversion process from the excitation absorbing  
326 ytterbium sensitizer ions [40,41].

327 It could be observed that regardless of the layering conditions the first bilayer of the as prepared  
328 coating resulted in an increase of up-conversion emission intensity. Thus, this bilayer was the most  
329 efficient in shielding the upconversion luminescence obtained from the core as the coating does  
330 not have optically active components. After the first bilayer the upconversion luminescence  
331 decreased with increasing number of layers below the core materials' luminescence (Figure S10).  
332 Crosslinking of the as prepared coating resulted in further enhancement of the luminescence of the  
333 first bilayer but with the increasing number of bilayers with the ionic concentrations of 0.1 and 0.2  
334 M NaCl prepared without additional fluoride the enhancement was lost. However, with the coating



335 prepared at the presence of additional fluoride the upconversion luminescence enhanced also after  
 336 crosslinking (Figure 6). This could indicate that the water in the bilayer structure could be  
 337 responsible for the luminescence emission quenching with the increasing number of bilayers. The  
 338 crosslinking of the bilayers can also increase compactness of the coating which can affect the  
 339 absorption of the bilayers creating less interference with the emission. This confirmed that the first  
 340 steps of disintegration of UCNPs have started during the surface modifications if they are made in  
 341 aqueous environments without additional fluoride. The crosslinking of the coating does not  
 342 enhance the obtained upconversion luminescence if the surface has already been covered with OH<sup>-</sup>  
 343 from the water molecules responsible for the quenching [13,42]. While the total luminescence  
 344 intensity is decreasing through increasing amount of bilayers the red-to-green ratio of the materials  
 345 is the lowest with the five bilayers of PAA/PAH suggesting that the layers are shielding especially  
 346 the green upconversion luminescence from the H<sub>2</sub>O molecules (Table S4) [21,40]. When the  
 347 coating was prepared at the presence of additional fluoride the red-to-green ratio remained similar  
 348 regardless of the number of coated bilayers. This also suggests that the core particle remains intact  
 349 during the coating procedure. However, further research and studies on the behavior of the  
 350 upconversion luminescence with this type of coatings is still needed but the result obtained here  
 351 seem promising.



353 Figure 6. Upconversion luminescence of the coated and crosslinked UCNPs prepared with  
354 additional 10 mM NaF present at the coating (left) and integrated intensity of the upconversion  
355 luminescence with coatings prepared with or without additional fluoride (right).

356

## 357 **CONCLUSIONS**

358 A successful layer-by-layer coating of upconverting nanoparticles with PAA/PAH polyelectrolyte  
359 multilayers is demonstrated with crosslinking the bilayer structure. Our research provides new  
360 information about the use of the layer-by-layer method in protecting the upconverting  
361 nanoparticles from environmental effects. It also simultaneously introduces surface modifications  
362 offering the possibility for further biomolecule conjugation. We observed that using additional  
363 fluoride during the coating procedure the upconversion luminescence of the NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> core  
364 nanoparticles can be enhanced and maintained with increasing number of bilayers. Our research is  
365 in line with the previous observations that the disintegration is affected by the fluoride environment  
366 in aqueous solutions [15,39]. In contrast to the previous research, where disintegration is strong in  
367 the beginning of the measurement, our surface modifications shield the nanoparticles from  
368 disintegration especially during the first five hours.

369 The bilayer formation was confirmed with various methods and it was successful regardless of the  
370 ionic concentration used during the coating. The crosslinking of the PAA and PAH bilayers was  
371 successful and could be observed from both FT-IR spectra as well as from the zeta potential  
372 measurements. Crosslinking of the bilayers created more defined coating structure on the  
373 nanoparticle surface as the interlocked water molecules were evaporated during the procedure. As  
374 expected, the additive fluoride during the coating procedure had no effect on the bilayer formation  
375 but it had an effect on the upconversion luminescence intensity of the prepared materials.

376 The coatings successfully decelerated the fluoride release and disintegration of the upconverting  
377 NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> nanoparticles. Especially the crosslinked coating prepared with additional  
378 fluoride present had a slower fluoride release rate, suggesting that when the nanoparticle remains  
379 intact during the surface modifications the crosslinking of the bilayers decelerates the movement  
380 of fluoride ions into the aqueous solution. The decelerating effect of the crosslinking seems to be  
381 the most beneficial during the first five hours in water which makes it a good candidate for  
382 applications needing a short time shielding. In addition, all of the coatings are able to reduce the  
383 disintegration of the particles when immersed in aqueous solution compared with the  
384 disintegration of the non-coated nanoparticles.

385 Further research and focus are needed for the applicability of these surface modifications but the  
386 results presented here are promising. This invites new possibilities for further exploiting the  
387 various properties of the layer-by-layer method and the bilayer formation in the shielding of  
388 upconverting nanoparticles.

389

## 390 **SUPPLEMENTARY INFORMATION**

391 Additional characterization data for materials (FT-IR spectra, thermal analysis, zeta potential,  
392 fluoride release data, and upconversion luminescence spectra with red-to-green ratios) are  
393 available in the supporting information.

394

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## 401 REFERENCES

- 402 [1] Z. Farka, T. Juřík, D. Kovář, L. Trnková, P. Skládal, Nanoparticle-Based  
403 Immunochemical Biosensors and Assays : Recent Advances and Challenges, *Chem. Rev.*  
404 117 (2017) 9973–10042.
- 405 [2] R. Arppe, L. Mattsson, K. Korpi, S. Blom, Q. Wang, T. Riuttamäki, et al., Homogeneous  
406 Assay for Whole Blood Folate Using Photon Upconversion, *Anal. Chem.* 87 (2015) 1782–  
407 1788.
- 408 [3] M. Ylihärtilä, E. Harju, R. Arppe, L. Hattara, J. Hölsä, P. Saviranta, et al., Genotyping of  
409 clinically relevant human adenoviruses by array-in-well hybridization assay, *Clin.*  
410 *Microbiol. Infect.* 19 (2013) 551–557.
- 411 [4] G. Chen, H. Qiu, P.N. Prasad, X. Chen, Upconversion Nanoparticles: Design,  
412 Nanochemistry, and Applications in Theranostics, *Chem. Rev.* 114 (2014) 5161–5214.
- 413 [5] F. Auzel, Upconversion and Anti-Stokes Processes with f and d Ions in Solids, *Chem.*  
414 *Rev.* 104 (2004) 139–173.
- 415 [6] J. Zhou, Q. Liu, W. Feng, Y. Sun, F. Li, Upconversion Luminescent Materials : Advances  
416 and Applications, *Chem. Rev.* 115 (2015) 395–465.
- 417 [7] H.H. Gorris, U. Resch-Genger, Perspectives and challenges of photon-upconversion  
418 nanoparticles - Part II bioanalytical applications, *Anal. Bioanal. Chem.* 409 (2017) 5875–  
419 5890.
- 420 [8] T. Rinkel, A.N. Raj, S. Dühnen, M. Haase, Synthesis of 10 nm  $\beta$ -NaYF<sub>4</sub>:Yb,Er/NaYF<sub>4</sub>

- 421 Core / Shell Upconversion Nanocrystals with 5 nm Particle Cores, *Angew. Chemie - Int.*  
422 *Ed.* 55 (2016) 1164–1167.
- 423 [9] Y. Li, Y. Dong, T. Aidilibike, X. Liu, J. Guo, W. Qin, Growth phase diagram and  
424 upconversion luminescence properties of NaLuF<sub>4</sub>:Yb<sup>3+</sup>/Tm<sup>3+</sup>/Gd<sup>3+</sup> nanocrystals, *RSC*  
425 *Adv.* 7 (2017) 44531–44536.
- 426 [10] K.W. Krämer, D. Biner, G. Frei, H.U. Güdel, M.P. Hehlen, S.R. Lüthi, et al., Hexagonal  
427 Sodium Yttrium Fluoride Based Green and Blue Emitting Upconversion Phosphors,  
428 *Chem. Mater.* 16 (2004) 1244–1251.
- 429 [11] J.L. Sommerdjik, On the Excitation Mechanisms of the Infrared-Excited Visible  
430 Luminescence in Yb<sup>3+</sup>, Er<sup>3+</sup>-doped Fluorides, *J. Lumin.* 4 (1971) 441–449.
- 431 [12] Y. Hossan, A. Hor, Q. Luu, S.J. Smith, P.S. May, M.T. Berry, Explaining the Nanoscale  
432 Effect in the Upconversion Dynamics of β-NaYF<sub>4</sub>:Yb<sup>3+</sup>, Er<sup>3+</sup> Core and Core–Shell  
433 Nanocrystals, *J. Phys. Chem. C.* 121 (2017) 16592–16606.
- 434 [13] R. Arppe, I. Hyppänen, N. Perälä, R. Peltomaa, M. Kaiser, C. Würth, et al., Quenching of  
435 the upconversion luminescence of NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> and NaYF<sub>4</sub>:Yb<sup>3+</sup>,Tm<sup>3+</sup> nanophosphors  
436 by water: the role of the sensitizer Yb<sup>3+</sup> in non-radiative relaxation., *Nanoscale.* 7 (2015)  
437 11746–11757.
- 438 [14] N.J.J. Johnson, S. He, S. Diao, E.M. Chan, H. Dai, A. Almutairi, Direct Evidence for  
439 Coupled Surface and Concentration Quenching Dynamics in Lanthanide-doped  
440 Nanocrystals, *J. Am. Chem. Soc.* 139 (2017) 3275–3282.
- 441 [15] S. Lahtinen, A. Lyytikäinen, H. Pääkilä, E. Hömppi, N. Perälä, M. Lastusaari, et al.,  
442 Disintegration of Hexagonal NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> Upconverting Nanoparticles in Aqueous  
443 Media: The Role of Fluoride in Solubility Equilibrium, *J. Phys. Chem. C.* 121 (2017)

- 444 656–665.
- 445 [16] O. Plohl, M. Kraft, J. Kovac, B. Belec, M. Ponikvar-Svet, C. Würth, et al., Optically  
446 Detected Degradation of NaYF<sub>4</sub>:Yb,Tm-based Upconversion Nanoparticles in Phosphate  
447 Buffered Saline Solution, *Langmuir*. 33 (2017) 553–560.
- 448 [17] Y. Wang, L. Tu, J. Zhao, Y. Sun, X. Kong, H. Zhang, Upconversion Luminescence of  $\beta$ -  
449 NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup>@ $\beta$ -NaYF<sub>4</sub> Core/Shell Nanoparticles: Excitation Power Density and  
450 Surface Dependence, *J. Phys. Chem. C*. 113 (2009) 7164–7169.
- 451 [18] K. Prorok, A. Bednarkiewicz, Energy Migration Up-conversion of Tb<sup>3+</sup> in Yb<sup>3+</sup> and Nd<sup>3+</sup>  
452 Codoped Active-Core/Active-Shell Colloidal Nanoparticles, *Chem. Mater*. 28 (2016)  
453 2295–2300.
- 454 [19] Z. Li, L. Wang, Z. Wang, X. Liu, Y. Xiong, Modification of NaYF<sub>4</sub>:Yb,Er@SiO<sub>2</sub>  
455 Nanoparticles with Gold Nanocrystals for Tunable Green-to-Red Upconversion  
456 Emissions, *J. Phys. Chem. C*. 115 (2011) 3291–3296.
- 457 [20] X. Chen, D. Zhou, W. Xu, J. Zhu, G. Pan, Z. Yin, et al., Fabrication of Au-Ag  
458 nanocage@NaYF<sub>4</sub>@NaYF<sub>4</sub>:Yb,Er Core- Shell Hybrid and its Tunable Upconversion  
459 Enhancement, *Sci. Rep.* 7 (2017) 41079.
- 460 [21] S. Wilhelm, M. Kaiser, C. Würth, J. Heiland, C. Carrillo-Carrion, V. Muhr, et al., Water  
461 dispersible upconverting nanoparticles: effects of surface modification on their  
462 luminescence and colloidal stability, *Nanoscale*. 7 (2015) 1403–1410.
- 463 [22] A. Sedlmeier, H.H. Gorris, Surface modification and characterization of photon-  
464 upconverting nanoparticles for bioanalytical applications, *Chem. Soc. Rev.* 44 (2014)  
465 1526–1560.
- 466 [23] O. Plohl, S. Kralj, B. Majaron, E. Fröhlich, M. Ponikvar-svet, D. Makovec, et al.,

- 467 Amphiphilic coatings for the protection of upconverting nanoparticles against dissolution  
468 in aqueous media, *Dalt. Trans.* 46 (2017) 6975–6984.
- 469 [24] E. Palo, S. Lahtinen, H. Pääkilä, M. Salomäki, T. Soukka, M. Lastusaari, Effective  
470 Shielding of NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> Upconverting Nanoparticles in Aqueous Environments  
471 Using Layer-by-Layer Assembly, *Langmuir*. 34 (2018) 7759–7766.
- 472 [25] G. Decher, Fuzzy Nanoassemblies: Toward Layered Polymeric Multicomposites, *Science*  
473 277 (1997) 1232–1237.
- 474 [26] M.M. De Villiers, D.P. Otto, S.J. Strydom, Y.M. Lvov, Introduction to nanocoatings  
475 produced by layer-by-layer (LbL) self-assembly, *Adv. Drug Deliv. Rev.* 63 (2011) 701–  
476 715.
- 477 [27] X. Hong, J. Li, M. Wang, J. Xu, W. Guo, J. Li, et al., Fabrication of magnetic luminescent  
478 nanocomposites by a layer-by-layer self-assembly approach, *Chem. Mater.* 16 (2004)  
479 4022–4027.
- 480 [28] G. Schneider, G. Decher, Functional core/shell nanoparticles via layer-by-layer assembly.  
481 Investigation of the experimental parameters for controlling particle aggregation and for  
482 enhancing dispersion stability, *Langmuir*. 24 (2008) 1778–1789.
- 483 [29] E. Palo, M. Salomäki, M. Lastusaari, Surface modification of upconverting nanoparticles  
484 by layer-by-layer assembled polyelectrolytes and metal ions, *J. Colloid Interface Sci.* 508  
485 (2017) 137–144.
- 486 [30] A.M. Balachandra, J. Dai, M.L. Bruening, Enhancing the Anion-Transport Selectivity of  
487 Multilayer Polyelectrolyte Membranes by Templating with Cu<sup>2+</sup>, *Macromolecules*. 35  
488 (2002) 3171–3178.
- 489 [31] E. Palo, M. Tuomisto, I. Hyppänen, H.C. Swart, J. Hölsä, T. Soukka, et al., Highly

490 Uniform Up-Converting Nanoparticles: Why You Should Control Your Synthesis Even  
491 More, *J. Lumin.* 185 (2017) 125–131.

492 [32] N. Bogdan, F. Vetrone, G.A. Ozin, J.A. Capobianco, Synthesis of ligand-free colloiddally  
493 stable water dispersible brightly luminescent lanthanide-doped upconverting  
494 nanoparticles, *Nano Lett.* 11 (2011) 835–840.

495 [33] J.J. Harris, P.M. Derosé, M.L. Bruening, Synthesis of Passivating, Nylon-Like Coatings  
496 through Cross-Linking of Ultrathin Polyelectrolyte Films, *J. Am. Chem. Soc.* 121 (1999)  
497 1978–1979.

498 [34] M.F. Elahi, G. Guan, L. Wang, M.W. King, Influence of Layer-by-Layer Polyelectrolyte  
499 Deposition and EDC/NHS Activated Heparin Immobilization onto Silk Fibroin Fabric,  
500 *Materials (Basel)*. 7 (2014) 2956–2977.

501 [35] Y. Guo, W. Geng, J. Sun, Layer-by-Layer Deposition of Polyelectrolyte-Polyelectrolyte  
502 Complexes for Multilayer Film Fabrication, *Langmuir*. 25 (2009) 1004–1010.

503 [36] Y. Ma, J. Sun, J. Shen, Ion-Triggered Exfoliation of Layer-by-Layer Assembled  
504 Poly(acrylic acid)/Poly(allylamine hydrochloride) Films from Substrates: A Facile Way  
505 To Prepare Free-Standing Multilayer Films, *Chem. Mater.* 19 (2007) 5058–5062.

506 [37] E. Harju, I. Hyppänen, J. Hölsä, J. Kankare, M. Lahtinen, M. Lastusaari, et al.,  
507 Polymorphism of NaYF<sub>4</sub>:Yb<sup>3+</sup>,Er<sup>3+</sup> up-conversion luminescence materials, *Z. Krist. Proc.*  
508 1 (2011) 381–387.

509 [38] P. Chodanowski, S. Stoll, Polyelectrolyte Adsorption on Charged Particles in the Debye-  
510 Hueckel Approximation. A Monte Carlo Approach, *Macromolecules*. 34 (2001) 2320–  
511 2328.

512 [39] D. Lisjak, O. Plohl, J. Vidmar, B. Majaron, M. Ponikvar-svet, Dissolution Mechanism of



- 513 Upconverting  $\text{AYF}_4:\text{Yb,Tm}$  ( $\text{A} = \text{Na}$  or  $\text{K}$ ) Nanoparticles in Aqueous Media, *Langmuir*.  
514 32 (2016) 8222–8229.
- 515 [40] I. Hyppänen, N. Höysniemi, R. Arppe, M. Schäferling, T. Soukka, Environmental Impact  
516 on the Excitation Path of the Red Upconversion Emission of Nanocrystalline  
517  $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ , *J. Phys. Chem. C*. 121 (2017) 6924–6929.
- 518 [41] M.T. Berry, P.S. May, Disputed Mechanism for NIR-to-Red Upconversion Luminescence  
519 in  $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ , *J. Phys. Chem. A*. 119 (2015) 9805–9811.
- 520 [42] F.T. Rabouw, P.T. Prins, P. Villanueva-delgado, M. Castelijns, R.G. Geitenbeek, A.  
521 Meijerink, Quenching Pathways in  $\text{NaYF}_4:\text{Er}^{3+},\text{Yb}^{3+}$  Upconversion Nanocrystals, *ACS*  
522 *Nano*. 12 (2018) 4812–4823.

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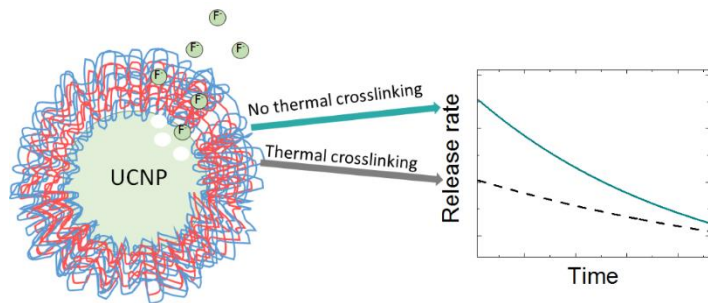
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536 **GRAPHICAL ABSTRACT**



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